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OPIATES INTERCEPT™ MICRO-PLATE EIA **for use with Intercept™ Drugs of Abuse (DOA) Oral Fluid Specimens**

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INTENDED USE

The OraSure Technologies, Inc. (OTI) Opiates Intercept™ MICRO-PLATE EIA is intended for use in the qualitative determination of opiates in oral fluid collected with the Intercept™ DOA Oral Specimen Collection Device. ***THIS TEST IS INTENDED FOR IN VITRO DIAGNOSTIC USE***

The OTI Opiates Intercept™ MICRO-PLATE EIA provides only a preliminary analytical test result. A more specific alternative chemical method should be used in order to obtain a confirmed analytical result. Gas chromatography/mass spectrometry (GC/MS) is the preferred confirmatory method.⁽¹⁾ Clinical consideration and professional judgment should be applied to any drugs of abuse test result, particularly when a preliminary, positive result is observed.

BACKGROUND

Opiates appear in saliva within one hour after use and, depending upon dose and rate of saliva flow, may persist in saliva for as long as 24 hours.^(2,3) Upon heroin administration, the major species detected in saliva is morphine and in urine it is conjugated morphine. However, morphine, codeine, and hydromorphone have been detected in saliva following administration.^(2,3) Heroin doses of 5 and 10 mg have been detected by immunoassay with high probability for 1 to 2 hours in saliva, while lower doses were not consistently detected at any sampling time.⁽³⁾ Conversely, 10 mg morphine is detectable in urine for up to 6 days after use, with the concentration approaching the NIDA cutoff of 300 ng/mL within about 36 hours.⁽²⁾

The OTI Opiates Intercept™ MICRO-PLATE EIA detects opiates including morphine, codeine, diacetylmorphine, hydrocodone, and hydromorphone.

PRINCIPLE OF THE ASSAY

The OTI Opiates Intercept™ MICRO-PLATE EIA is a competitive micro-plate immunoassay for the detection of opiates in oral fluid collected with the Intercept™ DOA Oral Specimen Collection Device. Specimen or standard is added to an EIA well in combination with an enzyme-labeled hapten derivative. In an EIA well containing an oral fluid specimen positive for opiates, there is a competition between the drug and the enzyme-labeled hapten to bind the antibody fixed onto the EIA well. EIA wells are then washed, substrate is added, and color is produced. The absorbance measured for each well at 450 nm is inversely proportional to the amount of opiates present in the specimen or calibrator/control. Because currently there are no SAMHSA assigned cutoffs for opiates testing using oral fluid, OTI recommends a cutoff of 10 ng/mL when testing oral fluid collected with the Intercept™ DOA Oral Specimen Collection Device. This cutoff is within the limit of detection by the OTI Opiates Intercept™ MICRO-PLATE EIA.

PRINCIPLE OF THE INTERCEPT™ DOA ORAL SPECIMEN COLLECTION DEVICE

Saliva is a complex mixture of parotid, submandibular, sublingual and minor salivary gland secretions mixed with mucin, bacteria, leukocytes, sloughed epithelial cells and gingival crevicular fluid. The fact that opiates are present in oral fluid following human use is well documented.⁽²⁾

The Intercept™ DOA Oral Specimen Collection Device was developed for the purpose of collecting oral fluid for diagnostic testing. The collection device consists of a treated absorbent cotton fiber pad affixed to a nylon stick (Collection Pad) and a preservative solution in a plastic container (Specimen Vial). The Collection Pad is impregnated with a mixture of common salts and gelatin which creates a hypertonic environment and an increased osmotic pressure wherever it contacts oral mucosal cells. The pad is placed in contact with the

gingival mucosa (between the lower gum and cheek) which enhances the flow of mucosal transudate across the mucosal surfaces onto the absorptive cotton fibers of the pad. Following the collection period, the Collection Pad is placed into a vial containing a preservative solution which serves to inhibit the growth of oral micro-organisms recovered on the Collection Pad. The vial is sealed with a plastic cap and transported to a laboratory for processing and testing. Following processing, a fluid containing a mixture of saliva components and the preservative solution is recovered which is suitable for testing for the presence of opiates in the OTI Opiates Intercept™ MICRO-PLATE EIA manufactured by OraSure Technologies, Bethlehem, PA. Refer to the Intercept™ DOA Oral Specimen Collection Device product insert for specific instructions on the proper collection, handling, and adequacy of oral fluid samples.

REAGENTS PROVIDED

KIT COMPONENTS	Catalog No. 1150IB	Catalog No. 1150IC
	480 Test Kit	9600 Test Kit
	Min. Qty.	Min. Qty.
Anti-Morphine Coated Plate -- Rabbit anti-morphine polyclonal antibody immobilized on a polystyrene plate supplied in dry form.	5	100
Enzyme Conjugate -- Horseradish peroxidase labeled with a morphine hapten diluted in a protein matrix of bovine serum with protein stabilizers.	60 mL	1 L
Substrate Reagent -- One bottle containing 3,3', 5,5' tetramethylbenzidine.	60 mL	1 L
Stopping Reagent -- Each bottle contains 2 N sulfuric acid.	60 mL	1 L
Oral Fluid Negative Calibrator – Oral Fluid Diluent negative for morphine.	2 mL	16 mL
Oral Fluid Negative Control – Oral Fluid Diluent containing 5 ng/mL (v/v) morphine.	2 mL	16 mL
Oral Fluid Cutoff Calibrator – Oral Fluid Diluent containing 10 ng/mL (v/v) morphine.	2 mL	16 mL
Oral Fluid Positive Control – Oral Fluid Diluent containing 20 ng/mL (v/v) morphine.	2 mL	16 mL

WARNINGS AND PRECAUTIONS

1. The handling of food or drink near the kit reagents is **NOT** recommended.
2. Proper handling of all reagents is strongly advised. It is suggested that disposable materials are used to avoid contamination of Substrate Reagent. Discard Substrate Reagent if obvious blue color develops.
3. Do **NOT** mouth pipet reagents. Handle all specimens and reagents as if potentially infectious.
4. Do **NOT** add sodium azide to samples as a preservative!
5. Keep all containers closed when not in use to avoid microbial contamination.
6. Do **NOT** use reagents past the expiration date.
7. Do **NOT** mix reagents from different kits or manufacturers.
8. Do **NOT** freeze reagents.
9. It is suggested that all OTI reagents be kept out of direct sunlight whenever possible.

STORAGE/STABILITY OF THE OTI OPIATES INTERCEPT™ MICRO-PLATE EIA

Store all reagents at 2-8°C until the expiration date on the kit label.

STORAGE/STABILITY OF THE INTERCEPT™ DOA ORAL SPECIMENS

Oral fluid specimens may be stored at 4°C (39°F) to 37°C (98°F) for a maximum of 21 days. Specimens must be tested in the OTI Opiates Intercept™ MICRO-PLATE EIA no later than 21 days following specimen collection, assuming that they have been maintained between 4°C and 37°C prior to testing. Specimens may be stored in either the original specimen storage vial or may be maintained as a processed fluid while being stored in a separate storage tube.

INTERCEPT™ DOA SPECIMEN PROCESSING PROCEDURE

MATERIALS REQUIRED BUT NOT PROVIDED

1. Tubes suitable for centrifuging Intercept™ DOA Specimen Vials.
2. Centrifuge capable of 600 - 800 x g

PROCEDURE (Refer to Intercept™ DOA Collector package insert for more information)

1. Record the specimen identification number from the Intercept™ DOA Specimen Vial.
2. Ensure that the specimen is within acceptable dating for testing, i.e. ≤ 21 days from the time of collection.
3. Hold the vial upright with the tip pointed up.
4. Move the pad away from the vial tip by gently tapping the vial.
5. Break the pointed tip of the vial off with your thumb.
6. Place a tube over the vial and invert the tube and vial.
7. Centrifuge at 600 - 800 x g for 15 minutes.
8. Assay or store the resulting eluate according to the procedures described herein.
9. A minimum of 0.7 mL of the eluate sample is required. This can be determined by centrifugation of the samples into graduated containers or by direct pipetting with a calibrated pipet adjusted to the specified volume.
10. If the minimum volume requirement is not met, a new sample should be collected. If this is not possible, a warning should accompany any data generated indicating that an insufficient amount of sample was collected and that this may affect the accuracy of the final result.

ASSAY PROCEDURE

MATERIALS REQUIRED BUT NOT PROVIDED

1. Semi-automated pipets (25, 50 and 100 microliters) with tips.
2. Micro-plate reader capable of reading at a dual wavelength of 450 and 630 nm.
3. Micro-plate washer.
4. Intercept™ DOA eluate sample(s) - 0.7 mL minimum.

PROCEDURE – Note: Allow all reagents and samples to come to room temperature (15-27°C) before use.

1. At the discretion of the operator, all samples, calibrators, and controls may be tested in duplicate. The inclusion of calibrators and controls is recommended in each run.
2. Add 25 microliters of sample, calibrator, or control to each well. Label wells appropriately.
3. Add 100 microliters of Enzyme Conjugate to each test well.
4. Incubate for 30 minutes at room temperature (15-27°C) in the dark.
5. Using a suitable plate washer, wash each well six (6) times with 300 microliters of distilled water.
6. Add 100 microliters of Substrate Reagent to each well and incubate 30 minutes at room temperature (15-27°C) in the dark.
7. Add 100 microliters of Stopping Reagent to each well.
8. Measure the absorbance at a dual wavelength of 450 nm and 630 nm within 15 minutes of stopping the reaction.

INTERPRETATION

Positive result: Any sample with an absorbance less than or equal to the Oral Fluid Cutoff Calibrator is considered a positive.

Negative result: Any sample with an absorbance greater than the Oral Fluid Cutoff Calibrator is considered a negative.

When interpreting duplicate results, the operator must be aware of several factors which may influence assay results. These include precise pipetting of specimens and reagents, effective washing of plates, and properly calibrated and maintained instrumentation. It is recommended that duplicate sample results which differ by more than 10% be retested. In addition, final pH levels of an oral fluid sample ≤ 5.0 may produce false positive results in the assay.

Also, negative patient results may indicate that either a patient specimen is negative for opiates, that the specimen contains less than 10 ng/mL opiates, or that an insufficient sample was collected.

QUALITY CONTROL

OTI provides negative and positive controls to monitor the daily performance of the OTI Opiates Intercept™ MICRO-PLATE EIA. The Oral Fluid Negative Control must have an absorbance greater than the Oral Fluid Cutoff Calibrator, while the Oral Fluid Positive Control must have an absorbance less than the Oral Fluid Cutoff Calibrator.

LIMITATIONS OF THE PROCEDURE

The assay is designed for use with oral fluid collected with the Intercept™ DOA Oral Specimen Collection Device. Other samples may produce variable results, and their use is not recommended.

SPECIFIC PERFORMANCE CHARACTERISTICS OF INTERCEPT™ DOA SPECIMENS

Analytical Sensitivity/Limit Of Detection - The Limit of Detection (LOD) was defined from the signal-to-noise ratio at the zero-drug concentration as the mean zero absorbance (A_0) minus the noise times three ($LOD = A_0 - 3SD$). The LOD was determined by obtaining the absorbance value for blank oral fluid devices and calculating the standard deviation (SD) of the absorbance values. The absorbance value minus 3SD was then extrapolated from the curve and represents the sensitivity of the assay. The LOD was calculated to be 1.4 ng/mL.

Precision - The precision of the OTI Opiates Intercept™ MICRO-PLATE EIA was assessed by testing Control Matrix containing 0, 5, 10, or 20 ng/mL morphine over a 5-day period. The intra-assay precision was calculated using 20 replicates of data from the first day. Inter-assay precision was calculated for the entire 5 days. The results of this testing are described in the following table:

MORPHINE (ng/mL)	INTRA-ASSAY % CV (n=20)	INTER-ASSAY % CV (n=20/day, 5 days)
0	3.6	7.5
5	6.4	8.9
10	6.6	9.5
20	6.9	8.7

Analytical Specificity/Cross-Reactivity - The analytical specificity of an immunoassay is defined as the cross-reactivity of substances in the assay which are structurally related to the target compound. The percent cross-reactivity of a compound in the OTI Opiates Intercept™ MICRO-PLATE EIA is defined as the apparent morphine concentration divided by the spiked concentration times 100.

The cross-reactivity of structurally related compounds was calculated at several spiked concentrations in Control Matrix. The following table indicates the apparent concentration of morphine for each substance at a concentration which cross-reacted in the assay.

The following compounds cross-react in the assay at the levels shown:

Compound	Tested Concentration (ng/mL)	Morphine Equivalents (ng/mL)	Cross-Reactivity (%)
6-Acetylmorphine	25	16	65
Codeine	1	4	>100
Dextromethorphan	10000	5	0.05
Diacetylmorphine	25	11	43
Dihydrocodeine	10	18.5	185
Hydrocodone	25	19	76
Hydromorphone	100	20	20
Levorphanol	1000	19	2
Meperidine	10000	8	0.08
Morphine	10	10	100
Morphine-3-β-Glucuronide	1000	19	2

Nalorphine	10000	16	0.16
Normorphine	1000	9	0.85
Oxycodone	1000	10	1.0
Oxymorphone	10000	10	0.10

The following compounds were spiked into Control Matrix at a target concentration of 10,000 ng/mL and tested for cross-reactivity. None were found to produce a signal less than or equal to that of the Cutoff Calibrator.

4-Aminophenyl Sulfone	Cocaethylene	L-Methamphetamine	Phenylephrine
Acetylsalicylic Acid	Cocaine	Lidocaine	Phenylpropanolamine
Alprazolam	Cotinine	Loperamide	Procainamide
Amobarbital	Cyclizine	Medazepam	Procaine
Ampicillin	D-Amphetamine	Methadone	Pseudoephedrine
Atropine	D-Methamphetamine	Metoprolol	Quinidine
β-Phenethylamine	Diphenhydramine	Naproxen	Salbutamol
Benzoylcegonine	Fenoprofen	Niacinamide	Temazepam
Butabarbital	Fluoxetine	Norchlordiazepoxide	Tolmetin
Butalbital	Gemfibrozil	Nordiazepam	Δ ⁹ -THC
Caffeine	Gentisic Acid	Nystatin	Theophylline
Chlordiazepoxide	Glipizide	PCP	Zomepirac
Chlorpromazine	Ibuprofen	Penicillin	
Clonazepam	Imipramine	Pentobarbital	
Clorazepate	L-Ephedrine	Phenobarbital	

It is possible that other substances and/or factors not listed above may interfere with the test and cause false results, e.g., technical or procedural errors.

ACCURACY

The clinical sensitivity and specificity of the OTI MICRO-PLATE assay were determined by testing specimens from chronic opiate users. A controlled dose study was also conducted to determine a minimum dose to produce a positive response in the OTI EIA. Both studies are described below. The cutoffs for EIA and GC/MS were 10 ng/mL and 15 ng/mL, respectively, for oral fluid specimens. For urine specimens, the EIA cutoff was 300 ng/mL and the GC/MS cutoff was 150 ng/mL (based on SAMHSA recommendations).

Study 1 - Chronic Opiates Users

Matched urine and oral fluid specimens were collected from ten (10) self-reported opiates users. An additional fifty (50) presumed negative samples were also included in the analysis. All sixty (60) samples were tested in the OTI Opiates Intercept™ MICRO-PLATE EIA and presumed positives (10) were also tested by gas chromatography/mass spectrometry (GC/MS). Results are presented in the following tables:

Result	Oral Fluid - OTI EIA*	Oral Fluid - GC/MS*
+	3	2
-	7	8

*Of the 10 samples presumed positive, the % sensitivity and % specificity of the OTI EIA as compared to GC/MS were 100.0% and 87.5%, respectively.

		Urine - OTI EIA (300 ng/mL Cutoff)	
		+	-
Oral Fluid - OTI EIA* (10 ng/mL Cutoff)	+	3	0
	-	4	53
% Relative Sensitivity = 42.9%		% Relative Specificity = 100.0%	

Result	Urine - OTI EIA	Urine - GC/MS**
+	7	8
-	3	2

**Of the 10 samples presumed positive, the % sensitivity and % specificity of the urine OTI EIA as compared to GC/MS were 87.5% and 100.0% respectively.

It is believed that the poor agreement between the OTI Opiates Intercept™ MICRO-PLATE EIA and the OTI Opiates MICRO-PLATE EIA (urine) is driven by differences in the metabolism of heroin and partitioning in oral fluid and urine. Opiates generally appear later in urine and persist for up to 3-6 days.⁽²⁾ Conversely, opiates in saliva is not detectable after 2-8 hours.⁽³⁾ These differences do not diminish the clinical utility of either fluid. However, users should be aware of these differences.

Study 2 - Controlled Dose Study

A total of 5 volunteers in a controlled environment were administered an initial 3 mg of intravenous heroin HCl, followed by a second 6 mg or 12 mg heroin dose 4 to 12 days later. Saliva and urine samples were collected before dosing and periodically for up to 74 hours after dosing. Saliva specimens (400 µL) were added to the collection pad of the Oral Fluid Collection Device and processed according to the package insert. Samples were then tested using the OTI assay and GC/MS.

Using a 10 ng/mL cutoff for oral fluid samples, opiates were detectable by EIA for up to 2 hours after a 12 mg heroin dose and not detectable at any time for a 3 or 6 mg dose. Of the GC/MS saliva results, only the initial post-12 mg dose samples contained drug concentrations above the 15 ng/mL LOD/LOQ. This demonstrates the ability of the OTI EIA to detect opiate concentrations at intravenous doses as low as 12 mg heroin HCl. The ability of the kit to detect opiates at doses between 6 mg-12 mg is unknown.

REFERENCES

1. "Urine Testing for Drugs of Abuse," National Institute on Drug Abuse (NIDA) Research Monograph 73, 1986.
2. Schramm, W., Smith, R.H., and Craig, P.A., "Drugs of Abuse in Saliva: A Review," *Journal of Analytical Toxicology*, 1992; 16:1-9.
3. Cone, E., "Testing Human Hair for Drugs of Abuse. I. Individual Dose and Time Profiles of Morphine and Codeine in Plasma, Saliva, Urine, and Beard Compared to Drug-Induced Effects on Pupils and Behavior," *Journal of Analytical Toxicology*, 1990; 14:1-7.
4. Tenovuo, Jorma O., editor, Human Saliva: Clinical Chemistry and Microbiology, Volume 1 (CRC Press, Boca Raton, FL) 1989.
5. Intercept™ Oral Specimen Collection Device, Package Insert. Manufactured by OraSure Technologies, Inc., Beaverton, OR 97008.

Note: *Adulteration of reagents, use of instruments without appropriate capabilities, or other failure to follow instructions as set forth in the labeling can affect performance characteristics and stated or implied label claims.*
